

Synthesis of pyrinodemins A and B. Assignment of the double bond position of pyrinodemin A

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Abstract—Condensation of aldehyde **3a** with hydroxylamine **4b** afforded nitrone **2**, which underwent an intramolecular cycloaddition to give **1b**, the proposed structure of pyrinodemin A. A similar condensation of aldehyde **3a** and hydroxylamine **4a** provided pyrinodemin A **(1a)**, which has the double bond one carbon further away from the isoxazolidine. An analogous sequence gave pyrinodemin B **(21)**. © 2001 Elsevier Science Ltd. All rights reserved.

Kobayashi and co-workers reported the isolation of the novel bis 3-alkylpyridine substituted *cis*-cyclopent[c]isoxazolidine alkaloid pyrinodemin A (1b) from the marine sponge *Amphimedon* sp. that shows potent cytoxicity to murine leukemia L1210 (IC₅₀ 0.058 μ g/mL) and KB epidermoid carcinoma cells (IC₅₀ 0.5 μ g/mL) in vitro. ^{1,2} The structure of pyrinodemin A was assigned by analysis of the ¹H, ¹³C, and 2D NMR and mass spectra. The position of the double bond was tentatively assigned based on the fragmentation pattern in the mass spectrum.

prepared by condensation of aldehyde **3a** and hydroxylamine **4b**, which can be formed from aldehyde **3b**, not **3a**. It seemed unlikely that both aldehydes **3a** and **3b** are intermediates in the biosynthesis of pyrinodemin A. This analysis suggested that the double bond of pyrinodemin A is not between carbons 16' and 17' as in **1b**, but rather between carbons 15' and 16' as in **1a**. Retrosynthetically, and biosynthetically, **1a** is a dimer that can be prepared by the condensation of aldehyde **3a** and hydroxylamine **4a**, which can be formed from **3a**. Analysis of the chemical shifts of the allylic carbons

Retrosynthetically, and presumably biosynthetically as well, **1b** can be prepared by the well-known intramolecular [3+2] dipolar cycloaddition of **2**.³ Nitrone **2** can be

provides further support for this analysis. Both allylic carbons of pyrinodemin A absorb at δ 27.1,¹ as expected for **1a**. Analysis of the expected substituent effects suggests that the γ -nitrogen should shield C-18' of **1b**, which should therefore absorb at δ 25. The potent cytotoxicity of pyrinodemin A prompted us to undertake the syntheses of both **1a** and **1b** to provide material for further biological evaluation and to determine the double bond position unambiguously.

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Isomerization of 3-octyn-1-ol (5a) with NaH in 1,3diaminopropane at 80°C overnight⁴ afforded 7-octyn-1ol (6a),5 which was coupled6 with 3-bromopyridine using (Ph₃P)₂PdCl₂ and CuCl to give 7a. Hydrogenation of 7a over PtO₂ afforded saturated alcohol 8a, which was converted to bromide 9a8 in conc. hydrobromic acid at reflux. Alkylation of LiC=C(CH₂)₄OTHP⁹ with **9a** in DMPU¹⁰ and THF provided 10a in 45% overall yield from **5a**. Hydrogenation of **10a** over lead-poisoned Pd on CaCO₃ and quinoline in the presence of 1-hexene to prevent over reduction¹¹ afforded 96% of *cis* alkene **11a**, which was hydrolyzed with 6 M hydrochloric acid in MeOH to give 12a. Swern oxidation afforded aldehyde 3a in 91% yield from 11a. Oxime 12a was prepared by reaction with NH₂OH·HCl and NaOAc in MeOH. 12,13 Reduction of 12a with NaBH₃CN in MeOH at pH 3-4 at -40 to -20°C afforded hydroxylamine 4a. 12,13 Condensation of 3a and 4a in toluene containing Na₂SO₄ at 0°C to form the nitrone and heating at reflux for 1 h to effect cycloaddition gave 1a¹⁴ in 75% yield from 3a and 4a. The stereochemistry of 1a was confirmed by NOEs between H-15, H-16, and H-20. This sequence provides 1a in 11 steps from 3-octyn-1-ol in 29% overall yield.

A similar five-step sequence starting with the conversion of 3-nonyn-1-ol (5b) to 8-nonyn-1-ol (6b)¹⁵ and concluding with alkylation of LiC≡C(CH₂)₃OTHP¹⁶ with 9b in DMPU¹⁰ and THF provided 10b in 47% overall yield from 5b. Hydrogenation, hydrolysis and Swern oxidation afforded 84% of 3b, which was converted to hydroxylamine 4b as described above for 4a. Condensation of 3a and 4b in toluene containing Na₂SO₄ at 0°C to form the nitrone and heating at reflux for 1 h to effect cycloaddition gave 1b¹⁴ in 79% yield from 3a and 4b. This sequence provides 1b in 11 steps from 3-octyn-1-ol and 3-nonyn-1-ol in 30% overall yield.

Spectral comparison indicates that pyrinodemin A is not 1b, and is probably 1a. The EI mass spectra of 1a and 1b are quite similar. Significant differences are: 1a m/z 258 (4), 231 (15), 124 (17); 1b m/z 258 (26), 231 (0), 124 (0). The EI mass spectrum of natural pyrinodemin A¹⁷ is similar to that of 1a with significant peaks at m/z 231 and 124, but not 258. The ¹H NMR spectra of 1a and 1b are again very similar except for the allylic methylene groups, which absorb at δ 2.07–1.96 (m, 4)

in **1a** and natural pyrinodemin A and at δ 2.17–2.04 (m, 2) and 2.01 (dt, 2, J=6, 6) in **1b**. The allylic carbons absorb at δ 27.1 and 27.2 in **1a**, while they both absorb at δ 27.1 in the natural product. In **1b**, they absorb at δ 27.0 for C-14′ and δ 24.9 for C-18′, as calculated, due to greater shielding by the γ -nitrogen than by a γ -carbon.

However, neither the 13 C NMR spectra of **1a** or **1b** match well with the published data for pyrinodemin A. For instance, the alkene carbons absorb at δ 129.6 and 129.9 in **1a**, at δ 129.3 and 130.3 in **1b**, and at δ 129.3 and 129.3 in the natural product. For **1a**, there are five absorptions between δ 27.0 and 28.5: C-13, 27.0; C-14′ and C-17′, 27.1 and 27.2; C-18′, 27.5; C-19′, 27.8. In the published data for pyrinodemin A there are only three absorptions in this region: δ 27.9, 27.1, and 27.1. However, examination¹⁷ of the 13 C NMR spectrum of the natural product shows peaks in this region that are smaller than expected, but do correspond well to those of **1a**, suggesting that this is the structure of the natural product.

Kobayashi and co-workers recently reported the isolation of additional cytotoxic pyridine natural products from Amphimedon sp including oxime 13a¹⁸ and pyrinodemin B (21), which differs from pyrinodemin A (1a) by the absence of a double bond and a one-carbon shorter side chain. Coupling of aldehyde 3a with hydroxylamine 20 should provide an expeditious route to pyrinodemin B (21). Alkylation¹⁹ of the lithium acetylide ethylenediamine complex with 13-bromo-1tridecanol (14) in DMSO containing NaI afforded 15,20 which underwent Pd-catalyzed coupling to give 48% of 16 from bromo alcohol 14. Hydrogenation afforded 90% of saturated alcohol 17, which was oxidized to aldehyde 18. Reaction with NH2OH·HCl and NaOAc provided 85% of oxime 19 from 17. Low temperature reduction of 19 with NaBH₂CN in MeOH at pH 3-4 gave 92% of hydroxylamine 20, which was condensed with aldehyde 3a in toluene at 0°C to reflux to give 81% of pyrinodemin B (21)¹⁴ in 26% overall yield from 14. The ¹H NMR spectrum of 21 is identical to that reported for the natural product. The ¹³C NMR spectrum is similar, except for the presence of peaks at δ 27.0, 27.3, and 28.1 in synthetic **21**, but only δ 27.9 in the natural product as with synthetic and natural pyrinodemin A.

Pyrinodemins **1a** and **1b** are moderately toxic to DU 145 prostate cancer cells (IC₅₀ 8 μ M) and HT-29 colon cancer cells (IC₅₀ 8 μ M).²¹

In conclusion, we have developed efficient syntheses of pyrinodemins A (1a) and B (21) that proceed in 26–30% overall yield. Analysis of the spectral data indicates that pyrinodemin A is definitely not 1b and is probably 1a.

Acknowledgements

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- 14. 1a: ¹H NMR 8.46–8.41 (m, 4), 7.48 (br d, 2, J=7.9), 7.20 (dd, 2, J=7.9, 4.9), 5.39-5.30 (m, 2), 4.05 (ddd, 1, J=6)6, 6), 3.50–3.42 (br, 1), 2.89–2.78 (m, 2), 2.64–2.60 (m, 1), 2.60 (t, 4, J=7.6), 2.07–1.96 (m, 4), 1.82–1.72 (br, 1), 1.71–1.50 (m, 9), 1.50–1.36 (m, 6), 1.36–1.24 (m, 20); ¹³C NMR 150.0 (2 C), 147.2 (2 C), 137.9 (2 C), 135.7 (2 C), 130.0, 129.6, 123.2 (2 C), 77.8 (br), 72.6 (br), 57.2 (br), 49.9, 34.2 (br), 33.0 (2 C), 31.1 (2 C), 29.7 (2 C), 29.4 (2 C), 29.33, 29.30, 29.2, 29.11, 29.08, 28.8, 27.8, 27.5, 27.18, 27.12, 27.0, 26.4, 26.3; IR (neat) 1574, 1464, 1422; MS m/z 575 (20), 574 (49), 365 (29), 355 (18), 327 (12), 301 (10), 299 (9), 289 (11), 288 (9), 287 (19), 286 (24), 285 (30), 270 (11), 258 (4), 244 (22), 231 (15), 220 (31), 204 (3), 190 (13), 176 (17), 162 (12), 148 (15), 146 (11), 136 (10), 134 (13), 124 (17), 120 (22), 106 (100), 93 (94); HRMS (DEI) calcd for $C_{38}H_{60}N_3O$ (MH⁺) 574.4736; found 574.4753. **1b**: ¹H NMR 8.46–8.41 (m, 4), 7.49 (br d, 2, J=7.9), 7.20 (dd, 2, J=7.9, 4.9), 5.41–5.30 (m, 2), 4.05 (ddd, 1, J=6, 6, 6), 3.50-3.42 (br, 1), 2.89-2.79 (m,2), 2.64-2.60 (m, 1), 2.60 (t, 4, J=7.6), 2.17-2.04 (m, 2), 2.01 (dt, 2, J=6, 6), 1.80-1.72 (br, 1), 1.71-1.52 (m, 10),1.50-1.38 (m, 3), 1.36-1.24 (m, 22); ¹³C NMR 149.9 (2 C), 147.1 (2 C), 137.91, 137.89, 135.7 (2 C), 130.3, 129.3, 123.2 (2 C), 77.8 (br), 72.6 (br), 56.8 (br), 49.9, 34.2 (br), 33.0 (2 C), 31.1 (2 C), 29.7 (2 C), 29.5 (2 C), 29.36, 29.35,
- 29.27, 29.24, 29.10, 29.06, 28.7, 28.0, 27.2, 27.0, 26.4, 26.3, 24.9; IR (neat) 1654, 1574, 1464, 1422;; MS m/z 575 (14), 574 (32), 365 (22), 355 (16), 327 (5), 301 (7), 299 (4), 289 (21), 288 (11), 287 (24), 286 (41), 285 (36), 270 (9), 258 (26), 244 (15), 231 (0), 220 (19), 204 (4), 190 (22), 176 (30), 162 (12), 148 (15), 146 (6), 136 (13), 134 (9), 124 (0), 120 (21), 106 (100), 93 (82); HRMS (DEI) calcd for $C_{38}H_{60}N_3O$ (MH⁺) 574.4736; found 574.4761. **21**: ^{1}H NMR 8.46–8.41 (m, 4), 7.49 (br d, 2, J=7.9), 7.20 (dd, 2, J=7.9, 4.9, 4.06 (ddd, 1, J=6, 6, 6), 3.50–3.42 (br, 1), 2.88-2.79 (m, 2), 2.64-2.60 (m, 1), 2.59 (t, 4, J=7.6), 1.83–1.72 (br, 1), 1.71–1.38 (m, 13), 1.36–1.22 (m, 28); ¹³C NMR 149.9 (2 C), 147.1 (2 C), 137.89, 137.85, 135.7 (2 C), 123.1 (2 C), 77.7 (br), 72.6 (br), 57.1 (br), 49.9, 34.2 (br), 32.9 (2 C), 31.1 (2 C), 29.7, 29.5–29.6 (5 C), 29.48, 29.35 (2 C), 29.27, 29.09, 29.05, 28.7, 28.1, 27.3, 27.0, 26.4, 26.3; HRMS (DEI) calcd for C₃₇H₅₉N₃O (M⁺) 561.4658; found 561.4645.
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- 18. The spectral data for **13a** are identical to those reported for the natural product.
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- 21. We thank Dr. Bruce Littlefield, Eisai Research Institute for evaluating the cytotoxicity of **1a** and **1b**.